

PSdif-BA® Brown Adipocyte Differentiation Kit

Human brown adipocytes can now be derived from pluoripotent stem cells (ES or iPS cells) by activating the classic brown adiogenic pathways (Ref 1, 2). **PSdif-BA**® is designed for brown adipocyte derivation from human pluoripotent stem cells (ESC or iPSC) grown as a monolayer culture. The kit contains all necessary serum-free media formulated with polypeptide differentiation factors and modulators of the key adipogenic pathways in a 3-step procedure. It is optimized to use with the serum-free and feeder-free growth medium **PSGro**® (StemRD cat# PGro). Ref 1: Tseng, et al., Nature 454: 1000, 2008; Ref 2: Nishio, et al., Cell Metab. 16: 394, 2012

Package Size, Content and Storage

PSdif-BA® has two package sizes:

- Starter kit: catalog # PB-STR (or PB-000 for samples): 3 wells (6-well plate), ~5x10⁶ starting cells
- Regular kit: catalog # PB-REG: 15 wells (6-well plate), ~2.5x10⁷ starting cells

Components (5)	Intended Outcome	Starter Kit Size	Regular Kit Size
PSGro [®] Plus	ESC or iPSC preparation	30 mL	130 mL
PSdif-BA® A	mesoderm derivation	10 mL	50 mL
PSdif-BA® B	Hematopoietic precursor derivation	10 mL	50 mL
PSdif-BA® C	Brown adipocyte progenitor derivation	10 mL	50 mL
BAGro [®]	BA maturation & maintenance	55 mL	250 mL

Storage: 2 to 8°C. Keep from light. Do NOT freeze. Shelf Life: 1 month if stored as directed.

Other reagents required:

- 1. Matrigel™: for cell plating
- 2. Accutase or equivalent: for cell splitting
- 3. Phosphate Buffered Saline (PBS): for cell washing
- 4. ROCK inhibitor **Thiazovivin** (StemRD # Thia) or Y-27632 (# Y27632): for optimal cell plating
- 5. **PSGro**® (StemRD # PGro) or equivalent (e.g., mTeSR®): for hESC/iPSC maintenance
- 6. Oil Red O Solution (Electron Microscopy Sciences, cat# 26609): for oil droplet staining
- 7. PCR reagents for brown adipocyte markers (e.g., UCP-1 and PRDM16)

Cell Preparation in PSGro® Plus

Coating plates with Matrigel™: Refer to manufacturer's instruction.

Recovery of frozen cells in PSGro[®]: Refer to PSGro[®] User Manual on StemRD website for details. https://www.stemrd.com/index.php?/Media/PSGro-Medium/flypage.tpl.html

Adaptation of growing cells to PSGro®

Most human ESC or iPSC lines that have been cultured as feeder-dependent or feeder-independent culture can be adapted to PSGro[®]. Refer to PSGro[®] User Manual on StemRD website for details.

Cell plating in PSGro[®] Plus

- 1. Start from a routine culture of hESC or hiPSC in **PSGro**[®] or a similar serum-free medium (e.g., mTeSR[®]). Identify and remove differentiated cells by scraping and aspiration.
- 2. Aspirate the medium and rinse twice with PBS.
- 3. Add 0.5 mL of Accutase per well (6-well plate). Incubate at 37°C for 3 5 min and verify that colonies have become single cells or small clusters (2 10 cells/cluster) under microscope.
- 4. Add 2 mL/well **PSGro® Plus** and pipet up & down 2 3 times gently.
- 5. Transfer the detached cells to a conical tube. Centrifuge at 200 x g for 5 minutes at room temp.
- 6. Aspirate the supernatant. Resuspend pellet in 3 mL **PSGro**[®] **Plus** gently. *Note: adding Thiazovivin* (2.5 uM) or **Y-27632** (10 uM) to **PSGro**[®] **Plus** at this step markedly increases plating efficiency.

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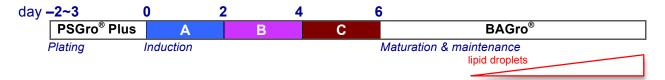


- 7. Plate the cells in a Matrigel™-coated well. For most cell lines, a 1:5 to 1:10 splitting from a routine, sub-confluent, culture may be appropriate while the ideal split may vary between lines.
- 8. Culture at 37°C, 5% CO₂ / 95% humidity. Refresh with **PSGro**[®] **Plus** (without ROCKi) daily.

Differentiation with PSdif-BA®

Cell density at the onset of differentiation: cell density is critical to achieve optimal brown adipocyte differentiation. The ideal cell density at the onset of induction is **40 - 60%** confluency. Less confluent culture may suffer from excessive cell loss upon induction whereas more confluent culture may result in over-confluency at the end of the procedure. Usually, if cells are plated in **PSGro® Plus** as recommended, they should reach 40 - 60% confluency in 2-3 days.

- **A.** Warm PSdif-BA® A to room temp. Aspirate PSGro® Plus, rinse once with PBS. Add 3 mL of PSdif-BA® A, incubate the cells at 37°C, 5% CO₂/95% humidity for **2** day.
 - Expected result: As a result of differentiation, cells become bigger and flatter.
- **B.** Warm PSdif-BA® B to room temp. Aspirate PSdif-BA® A. Add 3 mL PSdif-BA® B, incubate the cells for **2** days. Do not change the medium.
 - Expected result: Cell morphology continues to change. Some cell dislodging may occur, but the overall confluency increases as cells are becoming bigger and flatter.
- **C.** Warm PSdif-BA® C to room temp. Aspirate PSdif-BA® B. Add 3 mL PSdif-BA® C, incubate the cells for **2** days. Do not change the medium.
 - Expected result: Cells become oval or rounded in shape and some cell dislodging may continue.
- **D.** Warm **BAGro**[®] to room temp. Aspirate **PSdif-BA**[®] **C.** Add 2 mL **BAGro**[®], and incubate the cells for 2 3 days. Refresh the medium every 2 3 days.
 - Expected result: Cells shape stabilizes as rounded or oval. Lipid droplets typically appear after 4-8 day in **BAGro**[®] (day $8 \sim 12$ from induction) and increase over the next few days. Maturation and survival of the resulting brown adipocytes are expected over the next several weeks.



Trouble-shooting

- **1.** Excessive amount of cell dislodging or over-confluence: This usually occurs when cell density is too low or too high at the onset of induction. Start the induction at 40% to 60% confluence.
- **2.** Low efficiency: A large number of factors influence adiopogenic efficiency of a particular pluoripotent stem cell line. The main determinants include:
 - The quality of the starting cells: it is crucial to maintain cells at their full pluripotent state before induction.
 - The well-known diversity between pluoripoent cell lines, especially iPSC lines. Since ESC lines are generally more amenable to differentiation, a strongly-adiopogenic ESC line should be included as a control.
- 3. If no lipid droplet or brown adipocyte-specific gene expression appears after 2 weeks, the procedure has likely failed. As adiopogenic differentiation is governed by a large number of factors, many of which are out of StemRD's or operators' control, success cannot be guaranteed for all cell lines under every setting.

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